GVPC AGAR FOR *LEGIONELLA*

DETECTION AND ENUMERATION OF LEGIONELLA

BM07108

1 INTENDED USE

GVPC agar for *Legionella* is used for the enumeration, isolation and culture of *Legionella* species in water and other samples susceptible of harboring the bacteria.

2 HISTORY

In 1977, MacDade *et al.* Were the first to isolate the agent responsible for Legionnaire's Disease, a bacteria now known as *Legionella pneumophila*. After this discovery, numerous occurrences of *Legionella* isolation of *Legionella* were reported in fresh water environments such as water distribution systems, air conditioning, cooling towers, and spas. 48 species of *Legionella* are currently known.

In 1978, Weaver succeeded in cultivating Legionella on Mueller-Hinton chocolate agar. Feeley et~al., deduced that cysteine and ferric pyrophosphate could replace the vitamin and hemoglobin supplements found in the Mueller Hinton chocolate agar. Their work led to the formulation of a medium dubbed F-G agar. They determined as well that an atmosphere enriched at 2.5% CO2 was necessary for Legionella culture. In 1979, Feeley et~al. modified the the F-G medium by replacing acid hydrolysate of casein by yeast extract, and adding activated charcoal while eliminating starch. The resulting CYE media allowed better growth of Legionella. In 1980, Pasculle et~al. supplemented the CYE medium with ACES buffer. They demonstrated that this new medium, designated BCYE, offered a better recovery of Legionella and could be incubated aerobically. In 1981, Edelstein increased the sensitivity of the medium by adding α -cetoglutarate (BCYE α medium), and Wadowsky & Yee suggested incorporating glycine, vancomycin and polymyxin B (GVP medium) to obtain a selective culture media. In 1984, Dennis et~al. formulated the current GVPC medium by adding cycloheximide into GVP medium. They demonstrated that this selective medium allowed a greater level of Legionella isolation.

3 PRINCIPLES

Yeast extract constitutes a primary nutrient leading to *Legionella* growth.

Activated charcoal decomposes hydrogen peroxide (toxic metabolic by-product), captures the carbon dioxide and modifies the surface tension.

The ACES/KOH buffer maintains the pH and permits aerobic incubation.

Cysteine and ferric pyrophosphate represent indispensable nutritive elements for the growth of Legionella.

α-cetoglutarate is a growth activator for *Legionella*.

Secondary microflora are inhibited by the association of glycine, vancomycin, polymyxin B and cycloheximide.

4 INSTRUCTIONS FOR USE

Surface inoculation:

- To the surface of pre-poured plates (BM071), or to plates prepared as above, transfer 0.1 mL of the sample to be tested and its serial dilutions.
- Spread the inoculum with a sterile triangle or "hockey stick".
- Incubate at 36 ± 2 °C for 10 days.
- During the incubation period, examine the plates starting from the third day and three successive moments at intervals of 2 to 4 days.

After membrane filtration:

 Aseptically filter through a membrane a known volume of the sample to test (10 mL to 1000 mL of water sample).





✓ Inoculation :

On surface

✓ Incubation :

After membrane filtration

10 days at 36 ± 2°C

- Deposit the membrane on the surface of pre-poured plates (BM071), filtered side up and making sure that the membrane and agar are in close contact. The plates should be brought to room temperature before use.
- Incubate at 36 ± 2 °C for 10 days.
- During the incubation period, examine the plates starting from the third day and three successive moments at intervals of 2 to 4 days.

5 RESULTS

Colonies of *Legionella* spp. present a white to gray coloration. They can also have blue, pink, purple, maroon, greenish-yellow or dark red pigmentation that fades, becoming whiter and filamentous with age. Their surface is smooth with precise edges. Some strains may give a ground glass or "fried egg" aspect when observed through a binocular scope, while others may present a brilliant white fluorescence under a UV light.

Colonies of Legionella that develop on white membrane filters may have a different aspect to those that develop against a black background.

Enumerate each colony type separately. Select at least three characteristic colonies of *Legionella* on each of the agar plates. Re-streak each colony onto a plate of BCYE α without cysteine (BM073) and a plate of BCYE α (BM072) from the BT007 kit.

See ANNEX 1: PHOTO SUPPORT.

6 TYPICAL COMPOSITION

The typical composition can be adjusted to obtain optimal performance.

For 1 liter of medium:

- Yeast extract - Activated charcoal	•
- α-cetoglutarate, monopotassium salt	•
- ACES (2-[2-amino-2-oxoethyl)-amino] ethanesulfonic acid)	10.0 g
- Potassium hydroxide	
- L-cysteine, hydrochloride	
- Ferric pyrophosphate	
- Glycine	
- Vancomycin	1.0 mg
- Polymyxin B	80000 IŬ
- Cycloheximide	80.0 mg
- Bacteriological agar	12.0 g

pH of ready-to-use medium at 25° C : 6.9 ± 0.1 .

7 QUALITY CONTROL

Prepared media in plates: black agar, with visible particles of activated charcoal.

Typical cultural response after 5 days incubation at 36 °C (NF T 90-461) :

Microorganisms		Growth
Legionella pneumophila Enterococcus faecalis Escherichia coli	WDCM 00107 WDCM 00176 WDCM 00179	$66\% \leq R_2 \leq 150\%$ Inhibited Inhibited

8 STORAGE

Pre-poured media in Petri dishes : 2-8°C, shielded from light.

Expiration dates are indicated on the labels.

9 PRESENTATION

Pre-poured Petri dishes (&	90	mm)
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20 plates BM07108





10 BIBLIOGRAPHY

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11 ADDITIONAL INFORMATIONS

The information provided on the package take precedence over the formulations or instructions described in this document and are susceptible to modification at any time, without warning.

Document code : BM071/EN/2004-06 : 5.

Creation date : 2004-06 Update : 2013-06

Grounds for revision: General revision (§ 7: Quality control).



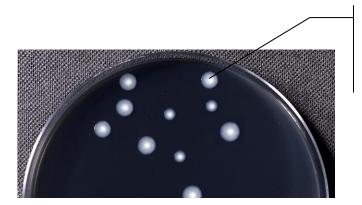


GVPC Agar for Legionella

Detection and enumeration of Legionella.

Reading:

Growth obtained after 10 days of incubation at 36 °C.



Legionella pneumophila

Characteristic colony
White to gray color with a smooth surface;
some colonies may present a ground glass
appearance under a binocular scope



